AMENDMENTS TO THE SPECIFICATION:

Please amend paragraph [0061] as follows:

[0061] DNA modifying enzymes and restriction endonucleases were used under standard conditions and in the buffers recommended by the manufacturers. General molecular cloning techniques and the electrophoresis of DNA and proteins were carried out according to standard procedures. L. lactis was transformed by electroporation of cells grown in the presence of glycin (Wells et al., 1993a). Plasmid DNA was routinely purified using the ~~Qiagen~~ QIAGEN® Plasmid Kit.

Please amend paragraph [0076] as follows:

[0076] Immunological testing of the sera showed that the treated mice did not show an immune response towards the expressed proteins. Serum was taken from the mice which were bled at day 8. This serum was analysed *via* Western blotting to check whether it contained antibodies against the proteins present in the medium fractions of the L. lactis cells. The medium fractions used were derived from the L. lactis strains MG1363 [pTREX1] and MG1363 [pT1mTFF1]. An equivalent of 1 ml of concentrated (phenol extraction and ethanol precipitation) medium fractions were analysed by SDS-polyacrylamide (20%) gel electrophoresis. After blotting to nitrocellulose filters, the filters were incubated for 1 hour with the serum solutions of the 4 groups of mice. The serum was diluted 500 times in 20ml nitrocellulose blocking buffer (~~Blotto~~ BLOTTO®: 100ml 10x PBS, 150ml 1M NaCl, 2ml TRITON X-100®, 25g fat-free milkpowder, water up to a total volume of 1 liter). As a secondary antibody, sheep anti-mouse IgG coupled to

horseradish peroxidase (HRP) was used. Using the 500 times diluted serum, no signal was detected.